

SAMPLING PROTOCOL FOR AMPHIBIAN CHYTRID FUNGUS (Bd and Bsal)

- Use fine tip swabs from Advantage Bundling/Medical Wire Co. (catalog number MW113). If these swabs are unavailable you can use other plastic handled swabs that you may have available. Avoid using wood handled swabs, as they can be associated with PCR inhibitors (and false-negative results).
 →You can also request a Sampling Kit from our laboratory. Write to AmphibianLab@sdzwa.org and send a mailing address and number of samples for supplies.
- 2. Using a single swab, gently swab the ventral surfaces of the skin approximately 20-30 times. Target areas to include the pelvic patch (5 passes with the swab), ventral thighs (5 passes each side with the swab) and toe webbing (5 passes on each foot). It is not necessary to swab the dorsal skin surfaces.
- 3. Break the swab approximately 2-3 cm from the tip and drop the tip into an empty screw-top tube. Avoid contact of the swab with the outside of the tube. Air-dry the swabs for approximately 5 minutes. Avoid contact of the drying swab with environmental surfaces, substrates, water droplets or other animals.
- 4. Using a fine tip permanent marker, label tube with:
 - Species (common name and/or genus)
 - Individual amphibian ID number
 - Date



 \rightarrow If you prefer to use a pre-printed label, make sure it is placed in the highlighted area of the tube.

5. Samples can be kept at room temperature for a week or so, but it is best to keep them cool and placed into a freezer (ultracold is ideal, but not absolutely necessary for short term storage). Most importantly, avoid high temperature extremes and direct sunlight.

Ship samples to:

Amphibian Disease Lab 15600 San Pasqual Valley Road Escondido, CA 92027 (760) 747-5471

If samples have been stored frozen, they should be shipped by overnight courier (Fed Ex, UPS, DHL) on dry ice.